TOTAL SYNTHESIS OF 4-ACETYLBENZOXAZOLIN-2-ONE

DAVID A. FIELDER*

Plant Research Centre

and F. WILLIAM COLLINS

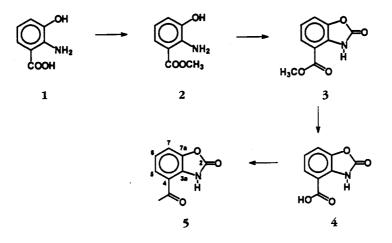
Centre for Food and Animal Research, Agriculture Canada, Ottawa, Ontario, Canada, K1A 0C6

ABSTRACT.—A practical synthesis of 4-acetylbenzoxazolin-2-one (4-ABOA) [5] in four steps and 57% overall yield, starting from 3-hydroxyanthranilic acid, is reported.

Recently, we reported the isolation and characterization of 4-acetylbenzoxazolin-2-one (4-ABOA, 5), a biosynthetically novel benzoxazolin-2-one (1) from the kernels of a hybrid line of Zea mays that exhibits resistance to Fusarium graminearum (2). Due to the complex isolation and the low concentration of 5 found in kernels, a synthesis was required to produce sufficient quantities for assessing its biological activity. Both the 5and 6-acyl synthetic analogues have been shown to possess analgesic, anti-inflammatory, and other pharmacological properties (3). Synthesis of these compounds follows a Friedel-Crafts acylation of benzoxazolin-2-one, which is commercially available (4). However, due to the para-directing nature of the aromatic ring substituents, this route was not possible for the synthesis of 5.

The three known naturally occurring benzoxazolin-2-ones (5-7) possess antifungal (8) and anti-inflammatory properties (9), as well as conveying resistance against certain insects (10). Synthesis of these compounds has thus far required the formation of a heterocyclic ring from a substituted phenol to produce the benzoxazolin-2-one. In the syntheses of benzoxazolin-2-one (BOA), 6methoxy- (MBOA), and 6,7-dimethoxybenzoxazolin-2-one (M₂BOA), the heterocyclic ring was formed via a Curtius rearrangement (11), or the reaction of urea (12), or phosgene (13) with the corresponding aminophenol.

Scheme 1 illustrates our synthetic sequence starting from 3-hydroxyanthranilic acid [1]. Prior to the formation of the heterocyclic ring, the reactive carboxylic acid was protected as the me-



SCHEME 1. Synthesis of 4-acetylbenzoxazolin-2-one (4-ABOA, 5) from 3hydroxyanthrinilic acid [1].

thyl ester [2] using H₂SO₄ in MeOH to prevent the formation of an isatoic anhydride or self-condensation product. The methyl ester 2 was treated with phosgene to form 4-carboxymethylbenzoxazolin-2-one [3] in high yield (96%). After quantitative hydrolysis of the ester, the free acid [4] was reduced to the methyl ketone with sequential treatment by methyllithium and trimethylchlorosilane (14) at room temperature to produce 5 (70%). The spectral data of synthetic 5 were identical with those previously reported (2). 4-Acetylbenzoxazolin-2-one [5] was formed from 3-hydroxyanthranilic acid in 57% overall yield.

EXPERIMENTAL

GENERAL EXPERIMENTAL PRCOEDURES.-Mps were determined on a Fisher-Johns melting point apparatus and are uncorrected. Uv spectra were taken on a Varian DMS200 spectrophotometer. Analytical tlc was performed on Si gel RP-18 (Whatman, 250 µm). Compounds were visualized by uv (365 nm) and with diethanolamine (5% w/v in EtOH). All moisture-sensitive reactions were conducted under a N2 atmosphere in ovendried glassware. Pyridine was distilled over CaH, and THF was distilled over sodium benzophenone ketyl prior to use. The sensitivity of compounds 1, 2, and 4 to light and air required all reactions to be performed in the dark and under a N2 atmosphere. Mass spectra were obtained on a Finnigan gc/ms system, model 4500, with a DS 400 data system. Compound 5 was injected on-column and chromatographed on a DB-5 fused silica capillary column (20 m \times 0.32 mm i.d.; 0.25 μ m film). The column was temperature programmed from 140 to 260° at 15°/min⁻¹ with He as the carrier gas at 10 psi. ¹H- and ¹³C-nmr spectra were run on a Bruker AM500 nmr spectrometer. Chemical shifts were referenced to MeOH- d_4 at 3.33 and 66.0 ppm for ¹H and ¹³C, respectively, and reported relative to TMS.

METHYL-3-HYDROXYANTHRANILATE [2].— A suspension of 3-hydroxyanthranilic acid [1] (1.53 g, 10.0 mmol) in MeOH (200 ml) was heated until the acid went into solution. Then, H_2SO_4 (40 ml, concentrated) was added dropwise over 3 min and the resulting solution refluxed for 1.5 h. The solution was cooled and neutralized with a saturated NaHCO₃ (aqueous) solution. The orange solution was extracted with EtOAc (3×500 ml). The combined extracts were dried over anhydrous MgSO₄ and the solvent removed *in vacuo*. Recrystallization from MeOH afforded pale orange crystals (1.42 g, 85% yield) of **2**: mp 95–97°; hrms m/z calcd for C₈H₉NO₃ 167.0582 ({M]⁺, 100), observed 167.0596, 135 (85), 107 (97), 79 (18), 52 (16); ¹H nmr δ 3.85 (COOCH₃), 5.8 (br, OH), 6.48 (dd, H-5), 6.80 (d, H-3), 7.46 (d, H-2); ¹³C nmr δ 51.6 (COOCH₃), 111.5 (C-1), 114.9 (C-5), 117.9 (C-4), 123.4 (C-6), 140.7 (C-2), 143.0 (C-3), 168.6 (COOCH₃).

4-CARBOMETHOXYBENZOXAZOLIN-2-ONE [3].—Phosgene(6 ml, 20% in toluene; 11.6 mmol) was added at room temperature over 10 min to a stirred solution of 2(1.22 g, 7.3 mmol) in pyridine (10 ml; dried over CaH₂). The mixture was allowed to stir overnight. The resulting pale orange solution was evaporated to dryness. The orange precipitate was dissolved in hot Me₂CO and filtered after being treated with decolorizing charcoal. After evaporation, the precipitate was recrystallized from Me₂CO/H₂O to afford 3 (1.36 g, 96% yield): mp 130–135° dec; hrms m/z calcd for $C_{9}H_{7}NO_{4}$ 193.0375 ([M]⁺, 70), observed 193.0372, 161 (100), 133 (12), 105 (62), 77 (29), 51 (20); ¹H nmr δ 3.98 (COOCH₃), 7.20 (dd, $J_{6,7}=J_{6,5}=8.1$ Hz, H-6), 7.45 (dd, $J_{5,6}=8.0$ Hz, $J_{5,7}=1.1$ Hz, H-5), 7.75 (dd, $J_{7,6}=8.2$ Hz, $J_{7,5}=1.1$ Hz, H-7); ¹³C nmr δ 156.7 (C-2), 132.9 (C-3a), 114.1 (C-4), 125.7 (C-5), 122.7 (C-6), 114.8 (C-7), 145.9 (C-7a), 166.5 (COOCH₃), 52.7 $(COOCH_3).$

4-CARBOXYBENZOXAZOLIN-2-ONE [4].—A mixture of **3** (402 mg, 2.1 mmol) and 0.5 N NaOH (30 ml) was stirred for 1 h, until a clear colorless solution remained. The solution was then acidified with 1 N HCl and extracted with EtOAc. The solvent was evaporated to dryness to give a quantitative yield of 4: hrms m/z calcd for C₈H₅NO₄ 179.0219, observed 179.0222, 161 (100), 105 (50), 77 (30), 51 (20); ¹H nmr δ 7.19 (dd, $J_{6,7}=J_{6,5}=8.1$ Hz, H-6), 7.42 (dd, $J_{5,6}=8.0$ Hz, $J_{5,7}=1.1$ Hz, H-5), 7.75 (dd, $J_{7,6}=8.2$ Hz, $J_{7,5}=1.1$ Hz, H-7); ¹³C nmr δ 156.7 (C-2), 133.0 (C-3a), 115.4 (C-4), 126.0 (C-5), 122.6 (C-6), 114.4 (C-7), 145.8 (C-7a), 167.8 (COOCH₃).

4-ACETYLBENZOXAZOLIN-2-ONE (4-ABOA, 5).—MeLi (10 ml, 1.4 M in toluene) was added rapidly to a stirred solution of 4 (50 mg, 0.28 mmol) in dry THF (5 ml) at 0° under N₂. The resulting solution was stirred for 2.5 h at room temperature and then quenched with freshly distilled (CH₃)₃SiCl (1000 µl). The solution was acidified with HCl (1 N, 3 ml) and stirred for 30 min. The solution was extracted with Et₂O, dried over MgSO₄, and dried *in vacuo*. The precipitate was taken up in *i*-PrOH-H₂O (1:1) and applied to a volume-calibrated column of Sephadex QAE (Pharmacia, Canada) in the hydroxide form and pre-equilibrated in *i*-PrOH-H₂O (1:1). Using this solvent, **5** eluted from the column in the first three bed volumes. Compound **5** was recrystallized from MeOH/H₂O to give plate-like crystals (35 mg, 70%); hrms m/z calcd for C₉H₂NO₃ 177.0426, observed 177.0465. Its uv and nmr spectra and mp were in complete agreement with those of the natural compound (2).

ACKNOWLEDGMENTS

The authors wish to thank Pierre Lafontaine and Neil Snider for the mass spectra and John Nikiforuk for the nmr spectra, and are also grateful for the assistance of Dr. O.E. Edwards. The cooperation of Ciba-Geigy Seeds (U.S. and Canada) and Dr. J. David Miller is appreciated. This work constitutes publication Nos. 1560 of the Plant Research Centre and 2244 of the Centre for Food and Animal Research.

LITERATURE CITED

- 1. H.M. Niemeyer, *Phytochemistry*, **27**, 3349 (1988).
- D.A. Fielder, F.W. Collins, B.A. Blackwell, C. Bensimon, and J.W. ApSimon, *Tetrabedron Lett.*, 35, 521 (1994).
- J.P. Bonte, D. Lesieur, and C. Lespagnol, Eur. J. Med. Chem., 9, 491 (1974).
- 4. H. Aichaoui, J.H. Poupaert, D. Lesieur,

and J.P. Hénichart, Tetrabedron Lett., 47, 6649 (1991).

- A.I. Virtanen and P.K. Hietala, Acta Chem. Scand., 9, 1453 (1955).
- A.I. Virtanen, P.K. Hietala, and O. Wahlroos, Arch. Biochem. Biophys., 69, 486 (1957).
- J.A. Klun, C.L. Tipton, J.F. Robinson, D.L. Ostrem, and M. Beroza, J. Agric. Food Chem., 18, 663 (1970).
- B.J. Long, G.M. Dunn, J.S. Bowman, and D.G. Rouley, Crop Sci., 17, 333 (1975).
- H. Otsuka, Y. Hirai, T. Nagao, and K. Yakasaki, J. Nat. Prod., 51, 74 (1988).
- V.H. Argandona, J.G. Luza, H.M. Niemeyer, and L.J. Corcuera, *Phytochemistry*, 19, 1665 (1980).
- F. Campos, J. Atkinson, J.T. Arnason, B.J.R. Philogene, P. Morand, N.H. Werstiuk, and G. Timmins, J. Chem. Ecol., 14, 989 (1988).
- E.H. Allen and S.K. Laird, J. Org. Chem., 36, 2004 (1977).
- 13. P.K. Hietala and O. Wahlroos, Acta Chem. Scand., 10, 1196 (1956).
- G.M. Rubottom and C. Kim, J. Org. Chem., 48, 1550 (1983).

Received 25 August 1994